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Document Title:	The Utility of Ultra High Performance Supercritical Fluid Chromatography for the Analysis of Seized Drugs: Application to
	Synthetic Cannabinoids and Bath Salts
Author(s):	Ira Saul Lurie
Document Number:	250512
Date Received:	August 2017
Award Number:	2014-R2-CX-K009

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Award Number: 2014-R2-CX-K009

The Utility of Ultra High Performance Supercritical Fluid Chromatography for the Analysis of Seized

Drugs, Application to Synthetic Cannabinoids and Bath Salts

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09/16/2016

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Project Period 01/01/2015 to 12/31/2016

Final Summary Overview

Final Summary Overview, NIJ award 2014-R2-CX-K009

The Utility of Ultra High Performance Supercritical Fluid Chromatography for the Analysis of Seized Drugs, Application to Synthetic Cannabinoids and Bath Salts

Purpose of Project

The purpose of this project is to investigate the role of ultra high performance supercritical fluid chromatography (UHPSFC) as a separation technique for forensic drug analysis. For this reason the challenging separation of emerging drugs such as synthetic cannabinoids and bath salts will be investigated. Emerging drugs contain similar solutes such as analogues, homologues, positional isomers, and stereoisomers. The latter two classes of compounds can present particularly difficult separation challenges for which UHPSFC appears well suited.

An additional goal of this study is to establish UHPSFC as a viable separation technique (recognized as a Category B test by SWGDRUG) comparable to already established techniques for the separation of emerging drugs. For UHPSFC, validated methods for the determination of synthetic cannabinoids and synthetic cathinones (bath salts) in seized exhibits using UV-PDA detection and single-quad MS detection will be developed.

Project Design

Experiments were designed to answer the question whether UHPSFC is a viable technique for forensic drug analysis, and whether as expected it is particularly well suited, compared to conventional techniques, for the separation of closely related substances (analogues, homologues, positional isomers, and diastereomers), which are present in emerging drug exhibits. The study consisted of three phases. For the first phase, using a UHPSFC instrument equipped with a PDA-UV detector and a single-quad mass

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spectrometric detector, optimum chromatographic and detector conditions were established for the separation of selected synthetic cannabinoids and substituted cathinones (Table 1 and Table 2).

Methods

A Waters Acquity UPC² system equipped with a PDA-UV detector and a QDA (single quad) MS detector was used for the UHPSFC experiments (Figure 1). The chromatographic conditions, including the effect of co-solvent, additive type, pressure, temperature, and column type (Table 3 and Table 4, Figure 2 and Figure 3) was investigated in order to determine the "optimum" separation conditions (most solutes resolved with resolution \geq 1) for the UHPSFC separation of synthetic cannabinoids and synthetic cathinones. The experimental steps taken are further summarized as follows:

For synthetic cannabinoids five minute gradient, plus one minute hold separations were carried out, so that the separation used the entire solvent space. UHPSFC separations with a CEL1 column and isopropanol modifier were further fined-tuned by varying gradient steepness (time of gradient), applied backpressure regulator (APBR), and temperature.

For synthetic cathinones five minute gradient, plus one minute hold separations or isocratic separations generally under six minutes were carried out, so that the separation used the entire solvent space. UHPSFC separations with a DIOL column and a methanol modifier were further fined-tuned by varying APBR and temperature.

Peak resolution was determined using standard resolution curves and the ratio of the valley to the height of the peak [1]. Principal component analysis (PCA) was performed on the UHPSFC, GC, and LC retention time data using IBM SPSS Statistics Version 21 (International Business Machines Corporation, Armonk, NY, USA). For PCA analysis the

data were autoscaled. Each variable was adjusted so that it had zero mean and unit standard deviation or variance.

For the synthetic cannabinoids and synthetic cathinones linearity, repeatability, and limits of detection were performed for both UV and MS detection using "optimized" UHPSFC conditions. Twenty simulated synthetic cannabinoid samples were prepared for analysis by adding known amounts of synthetic cannabinoids to ground leaf material (marshmallow, dogrose, beach bean, and honey weed). In addition, twenty simulated synthetic cathinone samples were prepared for analysis by adding known amounts of synthetic cathinones and an adulterant (lidocaine, benzocaine, caffeine, and pancake mix). See appendix S1 and S2 for sample preparation for both types of samples.

The use of two coupled columns in series with different selectivity to enhance the overall resolution of compounds was investigated. A mixture of 15 bath salts, as well as 10 sets of positional isomers of identical mass, were run on six individual columns (<10 minute runs) as well as eight coupled column combinations (Table 5 and Table 6). The Neue selectivity values [2], which determine the orthogonality of the individual columns, are investigated as a means to determine which coupled columns to use. For coupled column separation(s) further optimization was obtained by changing pressure.

Data Analysis

For the separation of synthetic cannabinoids and synthetic cathinones of primary interest was the separation of the positional isomers. These solutes are much more difficult to distinguish via retention time, and MS spectrum. Of secondary interest was the separation of a wide variety of controlled substances, which for the most part have different masses. In

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order to best evaluate UHPSFC as a separation technique, it was of interest to compare this chromatographic technique with established techniques such as GC and UHPLC.

For the various UHPSFC systems studied, the CEL1 column with an isopropanol modifier by far gave the best separation of the controlled synthetic cannabinoid JWH-018 and 9 non-controlled positional isomers. The separation was further optimized by varying the gradient steepness parameter (varied time of gradient) and temperature. All ten isomers were resolved (resolution ≥ 1) at a temperature of 55 degrees (resolution ≥ 1), in contrast to reversed phase (RP) UHPLC and GC where at best 3 and 4 out of 10 were resolved, respectively (Figure 4 and Figure 5) [3]. Under the same chromatographic conditions, the separation of 11 out of 22 controlled substances, including the separation of the positional isomers JWH-019 and JWH-122, was achieved (resolution ≥ 1) in under 10 minutes with both UV-PDA and MS QDA detection (Figure 6). For the same set of compounds, GC resolved 18 solutes in a 24-minute temperature-programmed run and UHPLC resolved at best 15 solutes in a 13-minute gradient run, both with a resolution ≥ 1 (Figure 7) [3]. Not only does UHPSFC exhibit great utility for the separation of positional isomers, it is also excellent for the separation of diastereomers as evidenced by the ability of the above chromatographic system to distinguish CP 47, 497 from its diastereomer Epi CP 47, 497 and the CP 47, 497 C8 homologue from its diastereomer 3-Epi CP 47, 497 C8. Although only useful for possible intelligence value, the AMY1 column with a methanol modifier was able to resolve from each other the individual enantiomers of all of the above solutes (Figure 8). HU-210 is well resolved from its diastereomer using an AMY1 column with an isopropanol modifier (Figure 9), in contrast to GC [4] and HPLC [5] where co-elution occurred.

For synthetic cathinones, the best overall separation of ten sets of positional isomers of identical mass was obtained with a DIOL stationary phase, a methanol modifier, and an

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ammonium formate additive under isocratic conditions. Further optimization was accomplished by varying the temperature, whereby 28 out of 34 positional isomers (sum of solutes resolved in each of the ten sets) were separated (resolution ≥ 1). This was in contrast to GC, RP UHPLC, and hydrophilic interaction UHPLC where at best for these same solutes 22 [6], 27[7], and 7 [7] solutes were separated, respectively [6]. The separation of the [M-H]⁺ 192 positional isomers by UHPSFC, GC, and UHPLC (RPC and hydrophilic interaction chromatography (HILIC) is shown in Figure 10 and Figure 11, whereby six out of eight, three out of eight, one out of eight, and five out of eight, respectively, are resolved (resolution ≥ 1). A five-minute isocratic separation of 10 out of 15 (resolution ≥ 1) controlled synthetic cathinones, by both UV and MS detection employing the "optimized conditions" for the positional isomers is shown in Figure 12. For the same set of compounds, GC, RPC, and HILIC resolved 13 [6], 12 [7], and 5 [7] solutes, respectively for an, 11-minute temperature-programmed run (Figure 13), an 11-minute gradient run and a 9-minute isocratic run (Figure 14). Although not the focus of this study, chiral separation of the individual "bath salts" using a UHPSFC chiral stationary phase is possible, which could be of intelligence value.

Principal component analysis for the various chromatographic types for the separation of synthetic cannabinoids (Figure 15) and "bath salts" (Figure 16) reveals that UHPSFC systems stand by themselves in a diffuse group, orthogonal to GC and RP UHPLC performed on a C18 column. In addition, for bath salts, UHPSFC is orthogonal to hydrophilic-interaction UHPLC performed on a silica column.

Based on the relatively high resolving power for synthetic cannabinoids and bath salts (particularly for positional and stereoisomers) and orthogonality to GC and UHPLC, UHPSFC belongs in the compendium of methods used for the analysis of seized drugs. The commonly employed RPC with a C18 column gives poor resolution of positional isomers. In terms of the overall resolution of controlled drugs and the separation of positional isomers, GC in combination with UHPSFC is the method of choice to provide complementary separation methods.

The great utility of employing the complementary detection modes of PDA-UV and electrospray single quad MS for the detection of synthetic cannabinoids and bath salts is shown in Figure 5 and Figure 10, respectively. MS detection offers far superior selectivity over UV detection, especially for non-isobaric compounds. For these solutes, overlapping peaks could be resolved using extracted ion chromatograms from either the $[M+H]^+$ or $[M+H-H_2O]^+$ ions of the analytes of interest. UV and MS data for these solutes, representing synthetic cannabinoids and bath salts, are given in Table 7 and Table 8, respectively. For these analytes, UV and MS libraries were created. UV spectra are particularly useful for determining subclasses of drugs and in distinguishing between positional isomers, particularly when differences occur in benzenoid substitution. JWH-018 has a unique UV spectrum compared to the other positional isomers (Figure 17). For a given mass, most positional isomers of bath salts gave unique UV spectrum (Figure 18-27).

For the synthetic cannabinoids shown in Figure 6, linearity analysis was performed and limits of detection ascertained for both UV and MS detection using the "optimized" UHPSFC conditions previously described (Table 9). For UV detection, excellent linearity was obtained, over two orders of magnitude, with $0.9996 \ge R^2 \ge 1.0000$. For MS detection, less favorable linearity was obtained, well over one order of magnitude for most solutes, with $0.9936 \ge R^2 \ge 0.9998$. For most solutes, MS detection offered lower limits of detection than UV detection, ranging in value from approximately a half to an order of magnitude higher. Run-to-run precision was examined for retention time (UV detection) and peak area (UV and

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MS detection) for each of the selected synthetic cannabinoids at three concentrations representing low, moderate, and high linearity concentration ranges (Table 10). Excellent run-to-run precision $(0.01 \ge \% \text{RSD} \ge 0.32, 0.18 \ge \% \text{RSD} \ge 2.92, 0.17 \ge \% \text{RSD} \ge 2.97)$ was obtained for retention time, peak area UV, and peak area MS, respectively. Excellent day-today precision $(0.12 \ge \% \text{RSD} \ge 0.50)$ over a one-week period was obtained for retention time (Table 11).

For the bath salts shown in Figure 10, linearity analysis was performed, and limits of detection ascertained for both UV and MS detection using "optimized" UHPSFC conditions previously described (Table 12). For the most part MS detection affords at least a two-orderof- magnitude linearity range. In contrast, UV detection offers at least a one-order-ofmagnitude linearity range. MS detection offers lower correlation coefficients than UV detection. For MS detection $0.9900 \ge R^2 \ge 0.9992$, while for UV detection $0.9994 \ge R^2 \ge 1.0000$. In addition, MS detection afforded two to three-orders-of-magnitude lower limits of detection than UV detection. Run-to-run precision was examined for retention time (UV detection) and peak area (UV and MS detection) for each of the selected bath salts, at three concentrations representing low, moderate, and high linearity concentration ranges (Table 13). Excellent run-to-run precision $(0.01\geq\% RSD\geq0.55, 0.31\geq\% RSD\geq3.00)$, (3.55 for 4-MePPP at low linearity concentration), $(0.31 \ge \% RSD \ge 2.86)$ was obtained for retention time, peak area UV, and peak area MS, respectively. Good day-to-day retention time precision over a one-week period was obtained especially if one takes into account relative retention times ($0.53 \ge$ %RSD \geq 1.52) (Table 14).

Twenty simulated synthetic cannabinoid samples were prepared for analysis by adding known amounts of synthetic cannabinoids to ground leaf material (marshmallow, dogrose, beach bean, and honey weed). In addition, twenty simulated synthetic cathinone samples

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were prepared for analysis by mixing known amounts of synthetic cathinones and an adulterant (lidocaine, benzocaine, caffeine, and pancake mix). Quantitative results for both synthetic cannabinoids and bath salts are shown in Table 15 and Table 16, respectively. For peaks containing single components, quantitation by UV detection resulted in good overall agreement for experimentally determined values in samples versus target values. In general, UV quantitation is more accurate than MS detection. For co-eluting peaks, MS detection provided a good estimate of the quantity of target drug present.

Custom reports were created for both qualitative and quantitative analysis (see appendix S3 and S4 for sample reports). For qualitative analysis, the tentative identification of a synthetic cannabinoid or a bath salt is accomplished by a combination of UV spectra and/or retention time, and MS spectrum base peak. For a given peak the qualitative report includes the UV apex spectrum, peak purity (compares UV spectra across peak), UV library search, MS spectra at peak apex and across the peak, and MS library search.

Protocols were developed for analysts for the analysis of synthetic cannabinoids and synthetic cathinones (appendix S5 and S6). Included are instructions for sample preparation, chromatographic conditions, representative chromatograms, structures, UV data, MS data, linearity data, limits of detection, and Water's instrumental software (Empower 3 files are available from the PI on request).

The use of coupled columns (DIOL + 2-PIC) significantly increased the separation of bath salts, whereby 31out of 34 positional isomers and 15 out of 15 controlled synthetic cathinones were resolved (Figure 12, Figure 28, and Figure 29).

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Scholarly Products Produced or in Process

Breitenbach, S.; Rowe, W. F.; McCord, B.; Lurie, I.S*, Assessment of ultra high performance supercritical fluid chromatography as a separation technique for the analysis of seized drugs: Applicability to synthetic cannabinoids, Journal of Chromatography A, 2016, 1440, 201-211.

Rowe, W.F.; Marginean, I.; Carnes (Breitenbach), S.; Lurie, I.S.*, The role of diode array ultraviolet detection for the identification of synthetic cathinones separated by ultra high performance supercritical fluid chromatography. Drug Testing and Analysis, under review. O'Brien, S.; Carnes, S.; Andrew, E.; Rowe W.F.; McCord, B., Lurie, I.S.* A Comparison of ultra high performance supercritical fluid chromatography, ultra high performance liquid chromatography, and gas chromatography for the separation of synthetic cathinones. manuscript in preparation.

Implications for Criminal Justice Policy and Practice in the United States

The use of UHPSFC would greatly assist the criminal justice system in the adjudication of emerging drug cases. In particular, it would aid in determining which if any controlled substances are present in seized drugs, particularly in the area of emerging drugs.

The use of emerging drugs, which are synthesized to circumvent the controlled substances laws, has greatly increased over the last few years. New structurally-similar compounds, such as synthetic cannabinoids and bath salts, are created by slightly modifying the chemical structure of a controlled substance. For the analysis of these solutes for legal purposes, the desired analytical methodology should have the ability to distinguish between similar solutes (analogues, homologues, positional isomers, and diastereoisomers). In this vein, separation methods such as liquid chromatography (HPLC, UHPLC) and gas

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chromatography have played a major role in the analysis of synthetic cannabinoids and bath salts. As evidenced for the analysis of synthetic cannabinoids, this project has demonstrated that UHPSFC has the potential to be superior to UHPLC and/or GC for the analysis of very similar solutes such as positional isomers of synthetic cannabinoids and bath salts and diastereoisomers of synthetic cannabinoids. For the separation of a diverse group for each class of emerging groups, including mainly non- isomeric solutes, UHPSFC has been demonstrated to be orthogonal to GC and UHPLC.

Detection methods also play a major role in the screening and confirmation of these analytes. Single quad MS soft-ionization techniques and UV detection typically used in LC, also provide for UHPSFC molecular mass and complementary structural information. In fact, most positional isomers of bath salts gave unique UV spectrum. UV detection subsequent to UHPSFC separation has proven to be excellent for quantitation of seized drugs. For co-eluting compounds, MS detection can provide a good estimate of the individual solutes. Based on the precision and high resolving-power of UHPSFC for the separation of emerging drugs, such as synthetic cannabinoids and synthetic cathinones, including positional isomers and diastereomers, it is recommended UHPSFC be included in the compendium of techniques for drug analysis, i.e., a SWG Drug category B test.

Appendix

References

1. Snyder, L., A rapid approach to selecting the best experimental conditions for high-speed liquid column chromatography. Part I—estimating initial sample resolution and the final resolution required by a given problem, J.Chromatogr. Sci. 2016, 10, 200-212.

2. Neue, U.D.; O'Gara, J.E.; Mendez., A,. Selectivity in reversed-phase separations Influence of the stationary phase , Journal of Chromatography A, 2006, 1127, 161-174.

3. Marginean, I.; Rowe, W.F.; Lurie, I.S., The role of ultra high performance liquid chromatography with time of flight detection for the identification of synthetic cannabinoids in seized drug, Forensic Science Int., 2015, 249, 83-91.

 Logan, B.L.; Reinhold, L.E.; Xu, A.; Diamond, F.X., Identification of synthetic cannabinoids in herbal incense blends in the United States, J. Forensic Sci., 2012, 57, 1168-1180.

5. Ciolino, L.A., Quantification of synthetic cannabinoids in plant materials using high performance liquid chromatography with UV detection (validated method), J. Forensic Sci., 2015, 60, 1171-1181.

6. Vaught, C., personnel communication 2106.

7. Obrien, S,. personnel communication 2106.

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Tables

Table 1- Synthetic cannabinoids

Solute	Structure	Chemical Formula
		monoisotopic mass
UR-144	O H ₃ C CH ₃	C ₂₁ H ₂₉ NO
	H ₃ C	311.2
	CH _{3s}	
CP47,497	он	$C_{21}H_{34}O_2$
		318.3
Epi CP 47, 497	ОН	C ₂₁ H ₃₄ O ₂
		318.3
RCS-4		C ₂₁ H ₂₃ NO ₂
	o h	321.2
JWH-073		C ₂₃ H ₂₁ NO
		327.2
XLR-11		C ₂₁ H ₂₈ FNO
	O F	329.2
CP47, 497 C8 homologue	OH OH	$C_{22}H_{36}O_{2}$
		332.3
	он	
3-ері СР47, 497 С8	ОН	$C_{22}H_{36}O_2$
Homologue		332.3
		a
JWH-250		$C_{22}H_{25}NO_2$
		335.2

JWH-203		C ₂₁ H ₂₂ CINO
		339.1
	CI	
JWH-018		C ₂₄ H ₂₃ NO
		341.2
JWH-016		C ₂₄ H ₂₃ NO
	0. N	341.2
JWH-018 2'-naphthyl isomer		C ₂₄ H ₂₃ NO
	o N	341.2
JWH-018 2'-naphthyl-N-(1, 1-		C ₂₄ H ₂₃ NO
		341.2
WH-018 2' nanhthyl N-(1, 2-		
dimethylpropyl) isomer		241 2
		341.2
JWH-018 2'-naphthyl-N-(2, 2-		C ₂₄ H ₂₃ NO
dimethylpropyl) isomer	o N	341.2
JWH-018 2'-naphthyl-N- (1		C ₂₄ H ₂₃ NO
ethylpropyl) isomer	0 N	341.2
methylbutyl) isomer		2/11 2
		541.2
	1	

JWH-018 2'-naphthyl-N-		C ₂₄ H ₂₃ NO
(2methylbutyl) isomer	o h	341.2
IWH-018 2'-naphthyl-N- (3		CadHaaNO
methylbutyl) isomer		241 2
	o u	541.2
JWH-019		C ₂₅ H ₂₅ NO
	0 N	355.2
JWH-122		C ₂₅ H ₂₅ NO
	0, N.	355.2
PB-22		$C_{23}H_{22}N_2O_2$
	0 N	358.2
AM-2201	$\langle \rangle$	C ₂₄ H ₂₂ FNO
	0 N F	359.2
АКВ-48		$C_{23}H_{31}N_{3}O$
	0 NNN	365.2
	H	
AB-Fubinaca	F	$C_{20}H_{21}FN_4O_2$
	0 NNN	368.2
	H NH2	
JWH-081		$C_{25}H_{25}NO_2$
		371.2
	1	l

RCS-8		$C_{25}H_{29}NO_2$
		375.2
JWH-200		$C_{25}H_{24}N_2O_2$
	o y no	384.2
HU-210	ОН	C ₂₅ H ₃₈ O ₃
	ОН	386.3
HU-211	OH	C ₂₅ H ₃₈ O ₃
	OH OH	386.3
	H 0	
AM- 694		$C_{20}H_{19}FINO$
		435.0
	~~~,	

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Table 2 - Bath Salts

Solute	Structure	Chemical Formula
		monoisotopic mass
Methcathinone	0 H U U	C ₁₀ H ₁₃ NO
		163.1
Nor-mephedrone	NH ₂	C ₁₀ H ₁₃ NO
		163.1
Mephredrone	O H	C ₁₁ H ₁₅ NO
		177.1
2-methylmethcathinone		C ₁₁ H ₁₅ NO
	O H N	177.1
3-methylmethcathinone		C ₁₁ H ₁₅ NO
		177.1
Buphedone	O H	C ₁₁ H ₁₅ NO
		177.1
Ethcathinone		C ₁₁ H ₁₅ NO
		177.1
N.N-dimethylcathinone	0	C ₁₁ H ₁₅ NO
	Ň,	177.1
4-fluoromethcathinone	° –	$C_{10}H_{12}FNO$
		181.1
3-fluoromethcathinone	0 	C10H12ENO
	F	181.1
Pentedrone	H H	C ₁₂ H ₁₇ NO
		191.1

Isopentedrone	H	C ₁₂ H ₁₇ NO
		191.1
4-methylethcathinone		C ₁₂ H ₁₇ NO
		191.1
2.2 dimethylmethylthingne	- н	
2,3-dimethymethcathinone		$C_{12}\Pi_{17}NO$
		191.1
2,4-dimethylmethcathinone		C ₁₂ H ₁₇ NO
		191.1
3,4-dimethylmethcathinone		C ₁₂ H ₁₇ NO
		191.1
2-ethylmethcathione	, ¹	C ₁₂ H ₁₇ NO
	Ň	191.1
4-methylbuphedrone		C ₁₂ H ₁₇ NO
		191.1
Methylone		$C_{11}H_{13}NO_3$
		207.1
2,3-methylenedioxymethcathinone		$C_{11}H_{13}NO_3$
		207.1
4-MePPP		C ₁₄ H ₁₉ NO
		217.1
3-IVIEPPP		$C_{14}\Pi_{19}NO$
		217.1
2-MePPP		C ₁₄ H ₁₉ NO
	N N	217.1
α-ΡΒΡ		C ₁₄ H ₁₉ NO
		217.1
Butylone		C ₁₂ H ₁₅ NO ₃
		221.1
	н	

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3,4-EDMC	о. Н 1	$C_{12}H_{15}NO_{3}$
		221.1
α-ΡVΡ		C ₁₅ H ₂₁ NO
		231.2
4-MePBP		$C_{15}H_{21}NO$
	· HCI	231.2
3-MePBP		$C_{15}H_{21}NO$
		231.2
2-MePBP		C ₁₅ H ₂₁ NO
		231.2
Pentylone		$C_{13}H_{17}NO_{3}$
		235.1
R-MMC		C ₁₃ H ₁₇ NO ₃
		235.1
MDPV		$C_{16}H_{21}NO_{3}$
		275.2
2,3-MDPV		C ₁₆ H ₂₁ NO ₃
		275.2
Naphyrone		C ₁₉ H ₂₃ NO
		281.2

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Table 3- UHPSFC experimental protocol for synthetic cannabinoids. Flow rate 1.25 mL/min, temperature 45 $^{\circ}$ C and APBR 2200 PSI.

Solutes	Columns	Modifiers	Detectors
22 controlled synthetic cannabinoids, JWH-018 and nine positional isomers	2-PIC DIOL DEA 1-AA	methanol	UV (PDA) MS (QDA)
	AMY1 CEL1 CEL2	methanol, acetonitrile, ethanol, isopropanol	

Table 4- UHPSFC experimental protocol for synthetic cathinones. Flow rate 1.25 mL/min, temperature  $45^{\circ}$ C and APBR 2200 PSI.

Solutes	Columns	Modifiers	Additives	Detectors
15 controlled bath salts and 34 positional isomers of	2-PIC DIOL DEA	methanol	10 mM ammonium formate, 10 mM	UV (PDA) MS (QDA)
controlled bath salts	AMY1 CEL1 CEL2	methanol, acetonitrile, ethanol, isopropanol	ammonium hydroxide	

Column	Particle Shape	Particle Size (µm)	Dimensions	Particle Technology
Torus 2-PIC	Spherical	1.7	3.0 x 100mm	BEH ^a
Torus DIOL	Spherical	1.7	3.0 x 100mm	BEH ^a
BEH 2-Ethylpyridine (2-EP)	Spherical	1.7	3.0 x 100mm	BEH ^a
Torus Diethylamine (DEA)	Spherical	1.7	3.0 x 100mm	BEH ^a
CSH Fluoro-Phenyl	Spherical	1.7	3.0 x 100mm	CSH ^b
HSS C ₁₈ Stable Bond	Spherical	1.8	3.0 x 100mm	HSS ^c

Table 5- Columns (3.0 x 100 mm) used in coupled column experiments

^aEthylene bridged hybrid

^bCharged surface hybrid

^cHigh strength silica

Table 6- UHPSFC experimental protocol for coupled columns. Flow rate 1.25 mL/min, temperature 40°C and APBR 2200 PSI.

Solutes	Single Column	Mobile Phase	Coupled Columns	Detectors
15 controlled bath salts and 34 positional isomers of controlled bath salts	2-PIC DIOL DEA 2-EP Fluoro-Phenyl C18	3% 10 mM ammonium formate in methanol, 97% carbon dioxide	DIOL + 2-PIC DIOL + Flouro-Phenyl DIOL + C18 2-PIC + DEA 2-PIC + C18 Flouro-Pheny I+ DEA DEA + C18 C18 + 2 EP	UV (PDA) MS (QDA)

Solute	Structure	UV _{MAX} (nm)	base peak [M-H] ⁺
HU-210		281	387.3
HU-211		281	387.3
CP47, 497	C C C C C C C C C C C C C C C C C C C	274	301.3 ^a
Ері СР47, 497	OH OH	274	301.3 ^a
CP47, 497 C8	QH C	274	315.3 ^a
3-epi CP47, 497 C8 homologue	OH OH OH	274	315.3 ^a
AKB-48	о н- ^N	266, 276, 300	366.2
XLR-11		215, 242, 296	330.2
UR-144		215, 242, 296	312.2
AB-Fubinaca		264, 270, 277, 300	369.2
AM-694		248, 311	436.1
RCS-4		211, 260, 312	322.2
RCS-8		212, 243, 299	376.2

Table 7 - Synthetic cannabinoids UV and MS data

JWH-250	< <u> </u>	212, 243, 299	336.2
	o N		
		242 200	240.2
J W H-203	<u>}</u>	245, 299	540.2
	o N		
DB 22		21/ 202	350.2
1 D-22		214, 272	557.2
	N		
IW/H_018		215 244 308	342.2
J WII-010	$\rangle = \langle$	213, 244, 300	542.2
JWH-018 2'-naphthyl-N-		213, 244, 316	342.2
(1, 2-dimethylpropyl) isomer	og / N		
	SI.		
JWH-018 2'-naphthyl-N-		213, 244, 316	342.2
(1 ethylpropyl) isomer	o N		
JWH-018 2'-naphthyl-N-		213, 244, 316	342.2
(1 methylbutyl) isomer	o N		
JWH-016		215, 244, 314	342.2
	o N		
WH 018 2' nonhthyl N		212 244 216	242.2
(1.1. dimethylenenyl) isomer		215, 244, 510	342.2
(1,1-dimethylpropyl) isomer			
IWH-018 2'-naphthyl-N-		213 244 316	342.2
(2 methylbutyl) isomer		213, 211, 510	512.2
(2 methyloutyl) isomer			
JWH-018 2'-naphthyl-N-		213, 244, 316	342.2
(2, 2-dimethylpropyl) isomer	0 N		
JWH-018 2'-naphthyl-N-	Č.	213, 244, 316	342.2
(3 methylbutyl) isomer	o the second sec		
	N N		
		1	

JWH-018 2'-naphthyl isomer		213, 244, 316	342.2
	0 N		
JWH-019		215, 244, 308	356.2
JWH-073		215, 244, 308	328.2
JWH-200		215, 244, 308	385.2
AM-2201		215, 244, 308	360.2
	o N P		
JWH-122		218, 244, 310	356.2
JWH-081		312	372.2

^a[M+H-H2O]⁺

Solute	Structure	UV _{MAX} (nm)	base peak [M-H] ⁺
α-ΡVΡ		238	232.2
2-MePBP		238, 278	232.2
3-MePBP		243, 284	232.2
4-MePBP	i n	249	232.2
α-PBP		238	218.2
3,4-MDPV		223, 269, 304	276.2
2,3-MDPV		222, 250, 324	276.2
Naphyrone		239, 280	282.2
4-MePPP		249	218.2
2-MePPP		238, 278	218.2
3-MePPP		243, 284	218.2
Pentedrone	N N N	239	192.1
Isopentedrone		262	192.1
4-methylethcathinone		250	192.1

# Table 8 - Synthetic cathinones UV and MS data

4-methylbuphedrone	O H - N	250	192.1
2-ethylmethcathinone	O H N	241, 280	192.1
2,3-dimethylmethcathinone		243, 281	192.1
2,4-dimethylmethcathinone	O H H	250	192.1
3,4-dimethylmethcathinone		254	192.1
Buphedone	O H N	239	178.1
Ethcathinone	O HN N	239	178.1
N, N-dimethylmethcathinone		238	178.1
4-fluoromethcathinone	F H	242	182.1
3-fluoromethcathinone		236, 282	182.1
Mephredrone	O H N	250	178.1
2-methylmethcathinone	O H Z	241, 281	178.1
3-methylmethcathinone	O H N	243, 286	178.1

Methcathinone		239	164.1
Nor-mephedrone	O NH ₂	249	164.1
Pentylone		224, 270, 304	236.1
R-MMC		228, 277, 304	236.1
Butylone	N U O	224, 270, 305	222.1
3,4 EDMC		226,270, 301	222.1
Methylone		224, 271, 304	208.1
2, 3- methylenedioxymethcathinone		222, 249, 326	208.1

Solute	Detection	Concentration	$\mathbf{R}^2$	Limit of
	Mode	Range µg/mL		Detection
				μg/mL
XLR-11	UV	0.230 - 59	1.0000	0.076
XLR-11	MS	0.230 - 7.4	0.9993	0.002
HU-210	UV	0.460 - 59	0.9999	0.153
HU-210	MS	0.230 - 14.8	0.9976	0.011
JWH-018	UV	0.244 - 62.5	1.0000	0.080
JWH-018	MS	0.244 - 7.8	0.9998	0.004
AKB-48	UV	0.230 - 59	0.9999	0.076
AKB-48	MS	0.057 - 7.4	0.9982	0.033
AM-2201	UV	0.115 – 59	0.9998	0.038
AM-2201	MS	0.115 - 7.4	0.9977	0.005
JWH-200	UV	0.976 - 62.5	0.9996	0.330
JWH-200	MS	0.122 - 7.8	0.9963	0.006
CP 47, 497 C8	UV	0.976 - 250	0.9998	0.330
CP 47, 497 C8	MS	0.488 - 31.2	0.9936	0.024
JWH-203	UV	0.390 - 50	0.9998	0.130
JWH-203	MS	0.097 - 6.25	0.9998	0.012
JWH-081	UV	0.390 - 50	0.9998	0.130
JWH -081	MS	0.097 - 6.2	0.9998	0.007
CP 47, 497	UV	0.976 - 250	0.9999	0.330
CP 47, 497	MS	0.244 - 7.812	0.9986	0.033
RCS-4	UV	0.390 - 50	0.9999	0.130
RCS-4	MS	0.097 - 6.2	0.9960	0.004
RCS-8	UV	0.390 - 50	0.9999	0.130
RCS-8	MS	0.097 - 6.25	0.9992	0.004
UR-144	UV	0.115 - 59	0.9999	0.038
UR-144	MS	0.115 - 14.8	0.9980	0.005
AB-Fubinaca	UV	0.460 - 59	1.0000	0.153
AB-Fubinaca	MS	0.057 - 7.4	0.9974	0.052
JWH-073	UV	0.488 - 62.5	0.9999	0.162
JWH-073	MS	0.122 - 15.6	0.9994	0.007
3-epi CP 47, 497 C8 (1) ^a	UV	1.952 - 250	0.9999	0.650
3-epi CP 47, 497 C8 (1) ^a	MS	0.488 - 31.2	0.9973	0.038
3-epi CP 47, 497 C8 (2) ^a	UV	1.952 - 250	0.9998	0.650
3-epi CP 47, 497 C8 (2) ^a	MS	0.488 - 31.2	0.9975	0.041
PB-22	UV	0.390 - 50	0.9999	0.130
PB-22	MS	0.097 - 6.2	0.9973	0.007
Epi CP 47, 497 (1) ^a	UV	1.952 - 250	0.9999	0.650
Epi CP 47, 497 (1) ^a	MS	0.488 - 31.2	0.9962	0.069
Epi CP 47, 497 (2) ^a	UV	1.952 - 250	0.99991	0.650
Epi CP 47, 497 (2) ^a	MS	0.488 - 31.2	0.9958	0.073
AM-694	UV	0.390 - 50	0.9999	0.130

Table 9 - Linearity and limits of detection for controlled synthetic cannabinoids

AM-694	MS	0.097 - 6.2	0.9996	0.007
JWH-122	UV	0.195 - 50	0.9999	0.065
JWH-122	MS	0.097 - 6.2	0.9988	0.003
JWH-019	UV	0.122 - 62.5	0.9997	0.041
JWH-019	MS	0.122 - 7.8	0.9996	0.004
JWH-250	UV	0.244 - 62.5	0.9998	0.081
JWH-250	MS	0.061 - 3.9	0.9974	0.002

^aindividual enantiomer

Table 10 - Run-to-run precision n=5 for synthetic cannabinoid solutions used for linearity studies.

Solute	Concentration	% RSD	% RSD	Peak Area
	ug/mL	RT	UV	MS
XLR-11	59.0	0.07	0.18	1.07
	3.68	0.05	0.79	1.50
	0.460	0.08	1.48	1.08
HU-210	59.0	0.16	0.16	1.48
	3.68	0.13	1.93	0.99
	0.460	0.20	1.39	1.46
JWH-018	62.5	0.04	0.14	0.37
	3.905	0.03	0.81	0.32
	0.448	0.05	1.76	1.17
AKB-48	59.0	0.10	0.61	1.28
	3.68	0.12	0.78	2.97
	0.460	0.06	0.75	2.24
AM-2201	59.0	0.10 0.54		0.56
	3.68	0.06 1.04		1.42
	0.460	0.04	1.63 0.8	
JWH-200	62.5	0.07	0.56 0.6	
	3.905	0.05	1.52	0.92
	0.448	0.03	2.92	0.85
CP 47, 497 C8	250	0.08	0.39	0.17
	15.62	0.14	0.87	1.93
	1.952	0.16	1.73 1.30	
JWH-203	50.0	0.03	0.41	0.79
	3.124	0.03	0.79	0.44
	0.390	0.16	0.65	1.60
JWH-081	50.0	0.03	0.38	0.89
	3.124	0.02	0.42	0.63

	0.300	0.02	0.53	1.05
CP 47 407	250	0.02	0.33	1.05
CI +/, +//	15.67	0.20	1.02	1.15
	1.052	0.18	1.02	1.07
DCS 4	50.0	0.23	0.28	0.76
KCS-4	2 124	0.03	0.28	0.70
	3.124	0.02	0.80	0.85
	0.390	0.09	2.32	0.81
RCS-8	50.0	0.02	0.20	0.08
	3.124	0.02	0.87	1.30
	0.390	0.09	2.79	1./3
UK-144	59.0	0.17	0.48	0.94
	3.68	0.08	0.48	2.11
	0.460	0.10	1.83	1./8
AB- Fubinaca	59.0	0.13	0.40	1.69
	3.68	0.06	1.10	2.86
	0.460	0.05	1.26	2.41
JWH-073	62.5	0.09	0.45	1.25
	3.905	0.01	1.16	0.54
	0.448	0.11	0.84	1.56
3-epi CP 47, 497 C8 (1)	250	0.09	0.30	1.36
	15.62	0.21	0.73	1.26
	1.952	0.16	1.78	1.50
3-epi CP 47, 497 C8 (2)	250	0.06	0.59	2.22
	15.62	0.14	1.24	2.08
	1.952	0.32	1.86	1.58
PB-22	50	0.03	0.69	1.30
	3.124	0.03	0.67	0.67
	0.390	0.05	0.54	1.21
Epi CP 47, 497 (1)	250	0.19	0.36	2.77
	15.62	0.21	0.72	2.21
	1.952	0.19	1.54	2.21
Epi CP 47, 497 (2)	250	0.16	0.45	1.89
	15.62	0.17	0.54	1.17
	1.952	0.21	2.08	1.89
AM-694	50	0.05	0.31	0.68
	3.124	0.08	0.51	1.12
	0.390	0.12	0.50	1.31
JWH-122	50	0.03	0.35	0.90
	3.124	0.08	0.52	1.26
	0.390	0.05	0.63	1.25
JWH-019	62.5	0.06	0.49	2.93
	3.905	0.02	0.51	1.09
	0.448	0.04	2.87	0.75
JWH-250	62.5	0.02	0.21	0.31
	3.905	0.03	0.65	0.51
	0.448	0.08	0.25	1.54

% RSD RT
0.13
0.31
0.22
0.22
0.29
0.50
0.27
0.26
0.12

Table 11- Day-to-day precision n = 5 for synthetic cannabinoid mix (50 µg/mL each solute)

Table 12 - Linearity and limits of detection for controlled bath salts

Solute	Detection	Concentration	$\mathbf{R}^2$	Limit of Detection
	Mode	Range µg/mL		μg/mL
α-PVP	UV	2.5 - 80	0.99980	0.094
α-PVP	MS	0.004 - 0.98	0.9970	0.00039
α-PBP	UV	2.5 - 80	0.99958	0.094
α-PBP	MS	0.004 - 2.96	0.9942	0.0012
MDPV	UV	0.625 - 80	0.99990	0.190
MDPV	MS	0.0013 - 0.98	0.9932	0.00039
Naphyrone	UV	0.625 - 80	0.99980	0.094
Naphyrone	MS	0.0013 - 2.96	0.9950	0.00039
4-MePPP	UV	0.3125 - 80	0.99995	0.094
4-MePPP	MS	0.004 - 2.96	0.9940	0.0012
4-methylethcathinone	UV	0.625 - 80	0.99994	0.094
4-methylethcathinone	MS	0.012 - 80	0.9992	0.0036
Pentedrone	UV	2.5 - 80	0.99940	0.378
Pentedrone	MS	0.036 - 26.6	0.9924	0.011
Buphedrone	UV	0.3125 - 80	0.99993	0.094
Buphedrone	MS	0.012 - 8.8	0.9987	0.0036
4-Fluoromethcathinine	UV	1.25 - 80	0.99980	0.378
4-Fluoromethcathinine	MS	0.004 - 0.98	0.9986	0.0012
3-Fluoromethcathinine	UV	2.5 - 80	0.99957	0.378
3-Fluoromethcathinine	MS	0.004 - 2.96	0.9966	0.0012
Mephedrone	UV	0.625 - 80	0.99992	0.190
Mephedrone	MS	0.012 - 0.98	0.9954	0.0036
Methcathinone	UV	2.5 - 80	0.99948	0.378
Methcathinone	MS	0.012 - 2.96	0.9916	0.0036
Pentylone	UV	1.25 - 80	0.99993	0.190
Pentylone	MS	0.004 - 8.8	0.9961	0.0012
Butylone	UV	0.625 - 80	0.99989	0.190
Butylone	MS	0.0013 - 0.98	0.9911	0.00039
Methylone	UV	1.25 - 80	0.99980	0.378
Methylone	MS	0.004 - 0.98	0.9900	0.0012

Coluto	Concentration ug/mL		% RSD	% RSD Peak Area	
Solute	UV	MS	RT	UV	MS
α-PVP	80	80	0.1	0.35	0.86
	5	0.98	0.05	1.65	2.76
	0.625	0.012	0.07	1.14	2.86
α-PBP	80	80	0.05	0.54	2.24
	5	0.98	0.06	0.55	2.48
	0.625	0.012	0.1	0.81	2.68
MDPV	80	80	0.02	0.31	0.97
	5	0.98	0.07	0.58	1.07
	0.625	0.012	0.09	3	1.1
Naphyrone	80	80	0.09	0.36	0.43
	5	0.98	0.09	0.96	2.73
	0.625	0.012	0.12	1.8	0.79
4-MePPP	80	80	0.09	1.8	0.52
	5	0.98	0.07	0.81	2.32
	0.625	0.012	0.55	3.55	0.95
4-methylethcathinone	80	80	0.01	0.38	0.91
	5	0.98	0.06	1.67	0.9
	0.625	0.012	0.04	1.55	2.11
Pentedrone	80	80	0.09	0.75	1.48
	5	0.98	0.1	2.68	2.41
	0.625	0.012	0.19	^a	1.94
Buphedrone	80	80	0.07	0.91	0.73
	5	0.98	0.09	1.52	2.4
	0.625	0.012	0.28	1.13	0.88
4-fluoromethcathinone	80	80	0.07	0.44	0.93
	5	0.98	0.06	2.21	1.78
	0.625	0.012	0.13	2.28	1.53
3-fluoromethcathinone	80	80	0.08	0.47	1.5
	5	0.98	0.09	2.57	1.28
	0.625	0.012	0.14	^a	2.03
Mephredrone	80	80	0.02	0.44	0.69
	5	0.98	0.03	0.99	0.31
	0.625	0.012	0.08	0.68	0.9
Methcathinone	80	80	0.06	0.68	1.23
	5	0.98	0.07	0.82	1.07
	0.625	0.012	0.11	^a	0.4

Table 13 - Run-to-run precision n=5 for bath salt solutions used for linearity studies

Pentylone	80	80	0.1	1.68	0.42
	5	0.98	0.06	1.33	2.08
	0.625	0.012	0.11	2.29	1.54
Butylone	80	80	0.05	2.43	1.06
	5	0.98	0.06	0.56	2.74
	0.625	0.012	0.14	1.81	0.93
Methylone	80	80	0.06	0.52	1.71
	5	0.98	0.03	0.85	1.24
	0.625	0.012	0.16	2.5	1.18

^ainsufficient signal to noise at concentration below linearity range

Table 14 - Day-to-day precision n= 5 for bath salt mix (50  $\mu$ g/mL each solute)

Solute	% RSD RT	%RSD RRT (Relative
	(UV detection)	to 4-
		Fluoromethcathinone)
α- PVP	1.74	1.516
MDPV	2.49	1.045
Pentedrone	2.08	0.980
4- Fluoromethcathinone	3.52	0.000
Pentylone	2.27	0.767
Mephedrone	3.86	0.887
Methylone	3.39	0.529

Table 15 - Anal	ysis of	simulated	synthetic	cannabinoid	samples
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mixture	mix 1					
plant	Marshmallow					
	UV MS					
compounds	CP 47,497	JWH-018	CP 47,497	JWH-018		
Target Concentration (ug/mL)	4	4	4	4		
Actual Concentration (ug/mL)		•		•		
sample 1	4.74	3.69	4.93	3.72		
sample 2	3.95	3.92	4.28	4.02		
AVERAGE	4.34	3.81	4.60	3.87		

mixture	mix 2					
plant	Bay Bean					
	U	V	MS			
compounds	JWH-018	JWH-081	JWH-018	JWH-081		
Target Concentration (ug/mL)	16	2	16	2		
Actual Concentration (ug/mL)						
sample 1	16.44	1.88	14.69	1.90		
sample 2	16.54	1.95	14.71	1.93		
AVERAGE	16.49	1.91	14.70	1.91		

mixture	mix 3					
plant	Marshmallow					
	UV MS					
compounds	RCS-4	JWH-019	JWH-122	RCS-4	JWH-019	JWH-122
Target Concentration (ug/mL)	2	2	2	2	2	2
Actual Concentration (ug/mL)						
sample 1	2.06	2.10	2.06	а	а	а
sample 2	2.07	2.10	1.95	а	а	а
AVERAGE	2.06	2.10	2.01			

mixture	mix 4					
plant	Marshmallow					
	UV MS					
compounds	JWH-018	AM-2201	JWH-018	AM-2201		
Target Concentration (ug/mL)	2	2 2		2		
Actual Concentration (ug/mL)						
sample 1	1.91	2.02	a	а		
sample 2	1.87	2.03	a	а		
AVERAGE	1.89	2.02				

mixture	mix 5					
plant	Marshmallow					
	UV MS					
compounds	RCS-8	JWH-081	RCS-8	JWH-081		
Target Concentration (ug/mL)	4	4	4	4		
Actual Concentration (ug/mL)						
sample 1	4.09	3.94	4.09	3.98		
sample 2	4.03	3.96	3.99	4.00		
AVERAGE	4.06	3.95	4.04	3.99		

mixture	mix 6					
plant	Bay Bean					
	UV MS					
compounds	JWH-019	JWH- 203	JWH-019	JWH-203		
Target Concentration (ug/mL)	4	2	4	2		
Actual Concentration (ug/mL)						
sample 1	b	b	4.11	1.93		
sample 2	b	b	4.02	1.94		
AVERAGE			4.07	1.93		

mixture	mix 7					
plant	Damiana					
	U	MS				
compounds	JWH-250	AKB-48	JWH-250	AKB-48		
Target Concentration (ug/mL)	2	2	2	2		
Actual Concentration (ug/mL)						
sample 1	1.88	1.94	1.96	2.00		
sample 2	1.86	2.06	1.93	2.08		
AVERAGE	1.87	2.00	1.94	2.04		

mixture	mix 8					
plant	Damiana					
	U	JV	MS			
compounds	HU-210	JWH-250	HU-210	JWH-250		
Target Concentration (ug/mL)	2	2	2	2		
Actual Concentration (ug/mL)						
sample 1	2.24	1.97	1.85	2.01		
sample 2	2.59	2.09	2.17	2.10		
AVERAGE	2.41	2.03	2.01	2.05		
mixture	mix 9					
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plant	Damiana					
	UV MS					
compounds	JWH-250	JWH-203	JWH-250	JWH-203		
Target Concentration (ug/mL)	2	8	2	8		
Actual Concentration (ug/mL)						
sample 1	b	b	2.32	8.55		
sample 2	b	b	1.87	7.66		
AVERAGE			2.09	8.11		

mixture					
		mix	. 10		
plant	Damiana				
	UV MS				
compounds	JWH-122	RCS-8	JWH-122	RCS-8	
Target Concentration (ug/mL)	16	8	16	8	
Actual Concentration (ug/mL)					
sample 1	15.40	7.07	13.56	6.75	
sample 2	14.75	8.03	13.02	7.53	
AVERAGE	15.08	7.55	13.29	7.14	

mixture	mix 11				
plant	Damiana				
	UV MS				
compounds	JWH73	JWH-018	JWH-073	JWH-018	
Target Concentration (ug/mL)	4	4	4	4	
Actual Concentration (ug/mL)					
sample 1	b	b	4.13	4.30	
sample 2	b	b	4.21	4.40	
AVERAGE			4.17	4.35	

mixture	mix 12				
plant	Bay Bean				
	UV MS				
compounds	AB-Fubinaca	JWH-122	AB-Fubinaca	JWH-122	
Target Concentration (ug/mL)	2	2	2	2	
Actual Concentration (ug/mL)					
sample 1	1.98	1.99	2.12	2.02	
sample 2	2.08	1.99	2.21	1.98	
AVERAGE	2.03	1.99	2.17	2.00	

mixture	mix 13					
plant			Ba	y Bean		
	UV MS					
compounds	RCS-4	JWH-073	JWH-81	RCS-4	JWH-073	JWH-081
Target Concentration (ug/mL)	8	16	4	8	16	4
Actual Concentration (ug/mL)						
sample 1	7.24	15.30	3.81	6.67	14.07	3.80
sample 2	7.55	15.06	3.86	6.71	13.76	3.705
AVERAGE	7.39	15.18	3.83	6.69	13.92	3.75

mixture	mix 14					
plant	Bay Bean					
	UV MS					
compounds	AB-Fubinaca	RCS-4	JWH-19	AB-Fubinaca	RCS-4	JWH-019
Target Concentration (ug/mL)	2	2	2	2	2	2
Actual Concentration (ug/mL)						
sample 1	1.96	1.89	1.99	2.06	1.96	1.99
sample 2	1.88	2.00	2.07	2.12	2.04	2.09
AVERAGE	1.92	1.95	2.03	2.09	2.00	2.04

mixture	mix 15				
plant	Honey Goat Weed				
	UV MS				
compounds	CP 47, 497	JWH-018	CP 47, 497	JWH-018	
Target Concentration (ug/mL)	2	2	2	2	
Actual Concentration (ug/mL)					
sample 1	1.97	2.30	1.99	2.33	
sample 2	1.85	2.14	1.97	2.25	
AVERAGE	1.91	2.22	1.98	2.29	

mixture	mix 16				
plant	Honey Goat Weed				
	UV MS				
compounds	AB-Fubinaca	AM 2201	AB-Fubinaca	AM 2201	
Target Concentration (ug/mL)	8	2	8	2	
Actual Concentration (ug/mL)					
sample 1	8.10	2.14	7.45	2.13	
sample 2	7.18	2.13	6.85	2.10	
AVERAGE	7.64	2.13	7.15	2.12	

mixture	mix 17				
plant	Honey Goat Weed				
	UV MS				
compounds	JWH-018	JWH-250	JWH-018	JWH-250	
Target Concentration (ug/mL)	2	4	2	4	
Actual Concentration (ug/mL)					
sample 1	b	b	2.07	4.05	
sample 2	b	b	2.08	4.58	
AVERAGE			2.07	4.31	

mixture	mix 18				
plant	Honey Goat Weed				
	UV MS				
compounds	JWH-203	JWH-122	JWH-203	JWH-122	
Target Concentration (ug/mL)	8	2	8	2	
Actual Concentration (ug/mL)					
sample 1	8.90	2.06	8.485	2.04	
sample 2	9.18	2.11	8.66	2.00	
AVERAGE	9.04	2.08	8.57	2.02	

mix 19				
Honey Goat Weed				
UV MS				
JWH-203	RCS-8	JWH-203	RCS-8	
2	2	2	2	
2.03	2.13	2.01	2.10	
1.96	1.99	1.97	2.00	
2.00	2.06	1.99	2.05	
	U JWH-203 2 2.03 1.96 <b>2.00</b>	mix           Honey G           UV           JWH-203         RCS-8           2         2           2         2           2.03         2.13           1.96         1.99           2.00         2.06	mix 19           Honey Goat Weed           UV         M           JWH-203         RCS-8         JWH-203           2         2         2           2         2         2           2         2         2           2.03         2.13         2.01           1.96         1.99         1.97           2.00         2.06         1.99	

mixture	mix 20				
plant	Bay Bean				
	UV MS				
compounds	AKB-48	JWH-073	AKB-48	JWH-073	
Target Concentration (ug/mL)	4	8	4	8	
Actual Concentration (ug/mL)					
sample 1	4.30	7.45	4.21	7.29	
sample 2	4.31	7.91	4.21	7.65	
AVERAGE	4.30	7.68	4.21	7.47	

^a no MS data available

^b no UV data due to coelution

#### Table 16 - Analysis of simulated synthetic cathinone samples

mixture	mix 1						
matrix	Lidocaine (1 mg/mL)						
	3-fluromet	hcathinone	Βι	ıtylone			
compounds	UV	MS	UV	MS			
Target Concentration (ug/mL)	75.00 75.00 75.00 75.00						
Actual Concentration (ug/mL)							
sample 1	72.08	74.39	72.53	74.28			
sample 2	70.93 74.72 71.24 74.48						
AVERAGE	71.50	74.56	71.89	74.38			

mixture	mix 2							
matrix		Pancake Mi	x (0.5 mg/mL	.)				
	4-methylet	hcathinone	α·	-PBP				
compounds	UV	MS	UV	MS				
Target Concentration (ug/mL)	37.50	37.50	37.50	37.50				
Actual Concentration (ug/mL)								
sample 1	37.25	34.67	40.02	36.33				
sample 2	37.18	35.66	38.72	36.78				
AVERAGE	37.21 35.16 39.37 36.55							

mixture	mix 3						
matrix		Lidocaine	(0.5 mg/mL)				
	Buphe	edrone	Pen	tedrone			
compounds	UV	MS	UV	MS			
Target Concentration (ug/mL)	37.50 37.50 37.50 37.						
Actual Concentration (ug/mL)							
sample 1	37.74	38.87	37.77	40.01			
sample 2	38.26	42.30	38.09	43.13			
AVERAGE	38.00	40.59	37.93	41.57			

mixture	mix 4						
matrix		Lidocaine	(0.5 mg/mL)				
	Naphyrone 4-methylethcathino						
compounds	UV	MS	UV	MS			
Target Concentration (ug/mL)	37.50	37.50	37.50	37.50			
Actual Concentration (ug/mL)							
sample 1	37.08	39.75	36.90	39.74			
sample 2	36.74	39.28	36.56	38.93			
AVERAGE	<b>36.91 39.51 36.73 39.3</b>						

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mixture	mix 5						
matrix		Benzocair	ne (1 mg/mL)				
	4-M	ePPP	Pen	tedrone			
compounds	UV	MS	UV	MS			
Target Concentration (ug/mL)	75.00 75.00 75.00 75.00						
Actual Concentration (ug/mL)							
sample 1	b	75.95	b	74.89			
sample 2	b	74.23	b	73.56			
AVERAGE		75.09		74.23			

mixture	mix 6							
matrix		Caffeine	(1 mg/mL)					
	Buty	lone	Met	hedrone				
compounds	UV	MS	UV	MS				
Target Concentration (ug/mL)	37.50	37.50	37.50	37.50				
Actual Concentration (ug/mL)								
sample 1	b	34.95	b	34.30				
sample 2	b	35.48	b	35.93				
AVERAGE	35.21 35.11							

mixture		mix 7						
matrix			Be	nzocaine (	187 ug/m	nL)		
	Methcar	thinone	Meth	ylone	Pent	ylone	α-Ρ	VP
compounds	UV	MS	UV	MS	UV	MS	UV	MS
Target Concentration (ug/mL)	62.50	62.50	62.50	62.50	62.50	62.50	62.50	62.50
Actual Concentration (ug/mL)								
sample 1	62.75	60.27	64.45	62.10	62.63	61.30	62.31	61.19
sample 2	62.17	59.72	63.30	59.79	61.90	59.58	61.91	59.98
AVERAGE	62.46	59.99	63.87	60.94	62.27	60.44	62.11	60.58

mixture		mix 8						
matrix			Pan	icake Mix	x (187 ug/1	nL)		
	Methcar	Methcathinone Methylone Pentylone α-PVI						VP
compounds	UV	MS	UV	MS	UV	MS	UV	MS
Target Concentration (ug/mL)	62.50	62.50	62.50	62.50	62.50	62.50	62.50	62.50
Actual Concentration (ug/mL)								
sample 1	65.76	65.98	65.49	65.38	66.45	64.61	65.28	64.39
sample 2	61.01	61.05	59.68	60.00	61.67	59.94	60.59	61.18
AVERAGE	63.38	63.52	62.58	62.69	64.06	62.27	62.94	62.78

mixture		mix 9						
matrix			Li	idocaine (	187 ug/mI	_)		
	Methcat	Methcathinone Methylone Pentylone α-PVP					VP	
compounds	UV	MS	UV	MS	UV	MS	UV	MS
Target Concentration (ug/mL)	62.50	62.50	62.50	62.50	62.50	62.50	62.50	62.50
Actual Concentration (ug/mL)								
sample 1	62.57	62.29	60.88	61.34	62.45	60.29	61.54	60.73
sample 2	62.85	61.75	61.97	60.77	62.12	61.44	61.80	60.55
AVERAGE	62.71	62.02	61.43	61.05	62.28	60.86	61.67	60.64

mixture	mix 10						
matrix	]	Pancake mix (	(187 ug/mL)				
	4-fluorometh	cathinone	4-Me	PPP			
compounds	UV	MS	UV	MS			
Target Concentration (ug/mL)	12.50	12.50	62.50				
Actual Concentration (ug/mL)							
sample 1	11.49	а	62.61	а			
sample 2	11.64	а	62.70	а			
AVERAGE	11.56		62.65				

mixture	mix 11						
matrix		Benzocaine (	187 ug/mL)				
	MD	PV	Penty	lone			
compounds	UV	MS	UV	MS			
Target Concentration (ug/mL)	50.00 50.00 25.00 2						
Actual Concentration (ug/mL)							
sample 1	49.87	52.17	25.42	26.05			
sample 2	49.11	50.59	25.16	25.75			
AVERAGE	49.49	51.38	25.29	25.90			

mixture		mix 12						
matrix			С	affeine (1	87 ug/mL	.)		
	Methcat	thinone	Methy	lone	Penty	ylone	α-P	VP
compounds	UV	MS	UV	MS	UV	MS	UV	MS
Target Concentration (ug/mL)	62.50	62.50	62.50	62.50	62.50	62.50	62.50	62.50
Actual Concentration (ug/mL)								
sample 1	63.07	61.36	63.74	61.63	62.70	60.53	64.40	60.72
sample 2	62.63	62.80	63.82	63.21	62.98	63.53	64.87	62.02
AVERAGE	62.85	62.08	63.78	62.42	62.84	62.03	64.63	61.37

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mixture	mix 13				
matrix	Benzocaine (187 ug/mL)				
	α- I	PVP	Butylone		
compounds	UV	MS	UV	MS	
Target Concentration (ug/mL)	25.00	25.00	75.00	75.00	
Actual Concentration (ug/mL)					
sample 1	24.89	24.16	74.33	72.42	
sample 2	24.10	23.37	72.37	71.25	
AVERAGE	24.50	23.76	73.35	71.83	

mixture	mix 14				
matrix		Pancake Mix	lix (187 ug/mL)		
	Pentedrone Mephedrone			edrone	
compounds	UV	MS	UV	MS	
Target Concentration (ug/mL)	62.50	62.50	62.50	62.50	
Actual Concentration (ug/mL)					
sample 1	63.71	62.365	63.64	62.05	
sample 2	62.46	59.79	62.67	61.13	
AVERAGE	63.08	61.08	63.15	61.59	

mixture	mix 15			
matrix	Caffeine (187 ug/mL)			
	Buphedrone Pentedrone			drone
compounds	UV	MS	UV	MS
Target Concentration (ug/mL)	25.00	25.00	50.00	50.00
Actual Concentration (ug/mL)				
sample 1	25.90	25.23	52.00	50.56
sample 2	25.04	25.29	50.29	50.44
AVERAGE	25.47	25.26	51.14	50.50

mixture	mix 16 Pancake Mix (187 ug/mL)				
matrix					
	MDPV Butyl			lone	
compounds	UV	MS	UV	MS	
Target Concentration (ug/mL)	62.50	62.50	31.25	31.25	
Actual Concentration (ug/mL)					
sample 1	62.98	63.95	31.98	31.97	
sample 2	63.64	64.23	32.05	31.16	
AVERAGE	63.31	64.09	32.01	31.56	

mixture	mix 17			
matrix	Lidocaine (75 ug/mL)			
	α- PBP		Buphedrone	
compounds	UV	MS	UV	MS
Target Concentration (ug/mL)	62.50	62.50	25.00	25.00
Actual Concentration (ug/mL)				
sample 1	69.14	64.12	25.78	25.35
sample 2	66.90	62.90	24.91	25.26
AVERAGE	68.02	63.51	25.34	25.30

mixture	mix 18				
matrix	Lidocaine (75 ug/mL)				
	4-methylethcathinone 4-MePPP			PPP	
compounds	UV	MS	UV	MS	
Target Concentration (ug/mL)	62.50	62.50	12.50	12.50	
Actual Concentration (ug/mL)					
sample 1	61.15	59.94	11.78	11.69	
sample 2	62.30	59.35	11.70	11.73	
AVERAGE	61.72	59.64	11.74	11.71	

mixture	mix 19				
matrix	Caffeine (75 ug/mL)				
	3-fluoromethcathinone Naphyrone			yrone	
compounds	UV	MS	UV	MS	
Target Concentration (ug/mL)	12.50	12.50	50.00	50.00	
Actual Concentration (ug/mL)					
sample 1	11.53	11.89	49.88	49.07	
sample 2	11.61	11.90	49.80	49.26	
AVERAGE	11.57	11.89	49.84	49.17	

mixture	mix 20 Caffeine (75 ug/mL)				
matrix					
	Mephedrone Pentedro			edrone	
compounds	UV	MS	UV	MS	
Target Concentration (ug/mL)	31.25	31.25	25.00	25.00	
Actual Concentration (ug/mL)					
sample 1	31.10	29.12	24.92	24.47	
sample 2	30.12	30.68	24.00	25.05	
AVERAGE	30.61	29.90	24.46	24.76	

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^a no MS data available ^b no UV data due to coelution

#### **Figures**



# Figure 1

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### UHPSFC Waters Achiral Columns 3.0 mm x 100 mm



and-Torus-Columns/nav.htm?cid=134696052&locale=en_US

Figure 2





Courtesy Waters

### Figure 3

UHPSFC JWH-018 Positional Isomers



- 1) JWH-016 m) JWH 018 2) 2'-napthyl isomer
- 4) 2'-napthyl-N-(1,2-dimethylpropyl) isomer
- 6) 2'-napthyl-N-(1 ethylpropyl) isomer
- 8) 2'-napthyl-N-(2 methylbutyl) isomer

- 3) 2'-napthyl-N-(1,1-dimethylpropyl) isomer
- 5) 2'-napthyl-N-(2,2-dimethylpropyl) isomer
- 7) 2'-napthyl-N-(1 methylbutyl) isomer
- 9) 2'-napthyl-N-(3 methylbutyl) isomer

Figure 4

### UHPLC VS GC JWH-018 Positional Isomers

Resolution ≥1



1) JWH-016m) JWH-0182) 2'-napthyl isomer3) 2'-napthyl-N-(1,1-dimethylpropyl) isomer4) 2'-napthyl-N-(1,2-dimethylpropyl) isomer5) 2'-napthyl-N-(2,2-dimethylpropyl) isomer5) 2'-napthyl-N-(2,2-dimethylpropyl) isomer6) 2'-napthyl-N-(1 ethylpropyl) isomer7) 2'-napthyl-N-(1 methylbutyl) isomer7) 2'-napthyl-N-(1 methylbutyl) isomer8) 2'-napthyl-N-(2 methylbutyl) isomer9) 2'-napthyl-N-(3 methylbutyl) isomer)

*See reference #3 for chromatographic conditions

## Figure 5

### UHPSFC Controlled Synthetic Cannabinoids



Figure 6

### UHPLC VS GC Synthetic Cannabinoids



*See reference #3 for chromatographic conditions.

### Figure 7



### Synthetic Cannabinoids Enantiomers

AM1 APBR 2200 PSI T = 45°C 15%-65% methanol 5 min. hold 1 min flow 1.25 ml/min.

## Figure 8





AMY1 APBR 2200 PSI T = 45°C 18%-53% isopropanol 5 min. hold 1 min flow 1.25 ml/min.

## Figure 9



### UHPSFC Pentedrone, 4-Methylethcathinone and Six Positional Isomers

- i) 4-methylethcathinone f) Pentedrone
- 3) 2-ethylmethcathinone

- 1) Isopentedrone
- 2) 4-methylbuphedrone

- 4) 2,3-dimethylmethcathinone
- 6) 3,4-dimethylmethcathinone

- 5) 2,4-dimethylmethcathinone
  - DIOL APBR 2200 PSI T = 40°C 3% methanol, 10mM ammonium formate flow 1.25 ml/min.

## Figure 10



Bath Salts Pentedrone, 4-methylethcathinone and Six Positional Isomers

- f) Pentedrone i) 4-methylethcathinone
- 3) 2-ethylmethcathinone
- 5) 2,4-dimethylmethcathinone

- 1) Isopentedrone 2) 4-methylbuphedrone
- 4) 2,3-dimethylmethcathinone
- 6) 3,4-dimethylmethcathinone

DIOL APBR 2200 PSI T = 40°C 3% methanol, 10mM ammonium formate flow 1.25 ml/min.

## Figure 11



### Figure 12

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### GC Bath Salts



a) α- PVPb) α- PBPc) MDPVd) Naphyronee) 4-MePPPf) Pentedroneg) Buphedroneh) 3-fluoromethcathinonehi) 3-fluoromethcathinoneii) 4-fluoromethcathinoneji) 4-fluoromethcathinonei) 4-methylethcathinonej) 4-fluoromethcathinoneji) 4-fluoromethcathinonen) Mephedrone o) Methylonek) Pentyloneki) Pentylonel) Methcathinonem) Butylonen) Mephedrone o) Methylone

Elite 5MS 30m x 250 x 0.25  $\mu$ m. Initial temp. 100°C for 1.0 min., ramp to 300°C at 20°C/min, hold final temp. for 2.0 min.

## Figure 13

### **UHPLC Bath Salts**

TOF-MS EIC's







a) α- PVPb) α- PBPc) MDPVd) Naphyronee) 4-MePPPf) Pentedroneg) Buphedroneh) 3-fluoromethcathinonehi) 3-fluoromethcathinoneii) 3-fluoromethcathinonei) 4-methylethcathinonej) 4-fluoromethcathinoneji) 4-fluoromethcathinonek) Pentyloneki) Pentylonel) Methcathinonem) Butylonen) Mephedroneo) Methylone

Figure 14

### Principal Component Analysis for Various Separation Techniques Synthetic Cannabinoids



The first factor accounts for 90.1% of the variance in the data while the second factor accounts for 7.9% of the variance

## Figure 15

### Principal Component Analysis for Various Separation Techniques Synthetic Cathinones



The first factor accounts for 78.3% of the variance in the data while the second factor accounts for 12.1% of the variance

Figure 16

### Normalized UV Spectra JWH018 Positional Isomers



- 1) JWH-016 m) JWH-018 2) 2'-napthyl isomer
- 4) 2'-napthyl-N-(1,2-dimethylpropyl) isomer
- 6) 2'-napthyl-N-(1 ethylpropyl) isomer
- 8) 2'-napthyl-N-(2 methylbutyl) isomer

- 3) 2'-napthyl-N-(1,1-dimethylpropyl) isomer
  5) 2'-napthyl-N-(2,2-dimethylpropyl) isomer
  7) 2'-napthyl-N-(1 methylbutyl) isomer
- 9) 2'-napthyl-N-(3 methylbutyl) isomer

## Figure 17



I) Methcathinone

11) Nor-methcathinone

Figure 18



Normalized UV Spectra Buphedrone, Mephedrone and Positional Isomers

Figure 19

### Normalized UV Spectra Pentedrone, 4-Methylethcathinone and Positional Isomers



f) Pentedronei) 4-methylethcathinonef1) Isopentedronei1) 4-methylbuphedronei2) 2-ethylmethcathinonei3) 2,3-dimethylmethcathinonei4) 2,4-dimethylmethcathinonei5) 3,4-dimethylmethcathinone

Figure 20



i) 4-fluoromethcathinone

j) 3-fluoromethcathinone

Figure 21



o) Methylone

o1) 2,3-methylenedioxymethcathinone

Figure 22



Normalized UV Spectra Alpha PBP, 4 MePPP and Positional Isomers

### b) alpha-PBP e) 4-MePPP e1) 2-MePPP e2) 3-MePPP

## Figure 23





n) Butylone

n1) 3,4-EDMC

Figure 24



Figure 25





m) Pentylone

m1) R-MMC

Figure 26



c) MDPV

c1) 2, 3-MDPV

Figure 27

#### Mix of 15 controlled cathinones with 2-PIC column



a)	α- PVP	b) α- PBP	c) MDPV	d) Naphyrone	e) 4-MePPP	f) Pentedrone
g)	Buphedrone	h) 3-fluorom	ethcathinone	i) 4- methylethc	athinone	j) 4-fluoromethcathinone
k)	Pentylone	I) Methcathi	none	m) Butylone	n) Mephedrone	o) Methylone

Figure 28
#### Mix of 15 controlled cathinones with DIOL + 2-PIC column



a) α- PVP b) α- PBP c) MDPV g) Buphedrone h) 3-fluoromethcathinone i) 4- methylethcathinone k) Pentylone I) Methcathinone

d) Naphyrone e) 4-MePPP m) Butylone

f) Pentedrone

j) 4-fluoromethcathinone

n) Mephedrone o) Methylone

Figure 29

**Supplemental** 

#### S1 - Sample Preparation for Synthetic Cannabinoids

The plant material (daminana, bay bean, marshmallow) was ground up into smaller components using sandpaper. The honey goat weed was already in powder form and did not need any further preparation.¹

Made a secondary stock solution (50 ug/ml in methanol) for each cannabinoid that had a concentration of 1 mg/ml in methanol or acetonitrile. Made a secondary stock solution (100 ug/ml in methanol) for each cannabinoid that had a concentration of 10 mg/ml. For the standard, 800 ul (50 ug/ml) or 400 ul (100 ug/ml) of the secondary stock solution of each drug was added in a 10ml volumetric flask and brought to volume in injection solvent (2:1:1 hexane: ethyl acetate: methanol) (4ug/ml).

For the samples, a 4mg/10ml or 10mg/25ml (plant material to final volume) ratio was used. The samples were prepared in the same way as the standard, but the amounts were adjusted depending on the concentration desired. The samples were brought to volume with injection solvent. The standard and samples were vortexed for 1 minute, filtered into LC vials, and run.

#### References

1. B. K. Logan, L. E. Reinhold, A. Xu, F. X., Diamond, Identification of synthetic cannabinoids in herbal incense blends in the United States , J. Forensic Sci. 57 (2012) 1168-1180.

#### S2 - Sample Preparation for the Synthetic Cathinones

A secondary stock solution containing the desired bath salts was prepared at a target concentration (75  $\mu$ g/mL, 62.5  $\mu$ g/mL, 50  $\mu$ g/mL, 37.5  $\mu$ g/mL, 31.25  $\mu$ g/mL, 25  $\mu$ g/mL) with methanol.

A standard and two duplicates were prepared by taking 1 mL of the secondary stock and placed into corresponding vials. 1 mg of adulterant (lidocaine, benzocaine, caffeine, or pancake mix) was added to the duplicates.

Alternatively, another standard and two duplicates were prepared by adding 250  $\mu$ L of the above secondary stock solution to corresponding vials. For the standard, 750  $\mu$ L of methanol was added to get a total volume of 1mL. For the duplicates, 750  $\mu$ L of methanol spiked with adulterant (2.5mg/10mL or 1mg/10mL) was added to each to get a total volume of 1mL.

To stay in linear range for the mass spectrometer, a 1:100 dilution was prepared for the standard and samples. All samples were vortexed, filtered into LC vials, and run.

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# Empower 3

## <u>83</u> screen cannabinoids

	SAMPLE	INFORMATIC	) N
Sample Name: Sample Type: Vial: Injection #: Injection Volume: Run Time:	mix 2 new std Standard 1:A,2 1 2.00 ul 11.0 Minutes	Acquired By: Sample Set Name: Acq. Method Set: Processing Method: Channel Name: Proc. Chnl. Descr.:	System sample 2 reprocess screen cannabinoids isopr CEL1 lurie syn can screen Extract 210.0 PDA Spectrum (210-350)nm
Date Acquired: Date Processed:	2/29/2016 3:44:07 PM EST 6/14/2016 11:14:31 AM EDT		





Peak #1 XIC - 5.134 - QDa Positive Scan: 342.15 m/z Peak #2 XIC - 9.552 - QDa Positive Scan: 372.22 m/z

#### PDA Result Table

	RT	Purity1 Angle	Purity1 Threshold	Match1 Spect. Name	Match1 Angle	Match1 Threshold	PDA/FLR Match2 Spect. Name	PDA/FLR Match2 Angle	PDA/FLR Match3 Spect. Name	PDA/FLR Match3 Angle	Base Peak (m/z)
1	5.134	0.656	1.442	JWH 18	0.529	1.835	JWH 73	0.534	JWH 19	0.535	342.15
2	9.552	0.681	1.793	JWH 081	0.171	2.249	JWH 200	7.595	AM 694	12.694	372.22

#### PDA Result Table

	MS Match1 Spect. Name	MS Match2 Spect. Name	MS Match3 Spect. Name
1	JWH 18	JWH 250	RCS4
2	JWH 81	RCS4	JWH 250



	SAMPLE	INFORMAT	ION
Sample Name: Sample Type: Vial: Injection #: Injection Volume: Run Time:	mix 2 new std Standard 1:A,2 1 2.00 ul 11.0 Minutes	Acquired By: Sample Set Name: Acq. Method Set: Processing Method: Channel Name:	System sample 2 reprocess screen cannabinoids isopr CEL1 uv quant 215 comp PDA Ch2 215nm@4.8nm -Compens.
Date Acquired: Date Processed:	2/29/2016 3:44:07 PM EST 6/14/2016 11:28:48 AM EDT		





	SAMPLE	<u>INFORMATIC</u>	) N
Sample Name: Sample Type: Vial: Injection #: Injection Volume: Run Time:	mix 2 new std Standard 1:A,2 1 2.00 ul 11.0 Minutes	Acquired By: Sample Set Name: Acq. Method Set: Processing Method: Channel Name: Proc. Chnl. Descr.:	System sample 2 reprocess screen cannabinoids isopr CEL1 ms quant 342 QDa Ch2 342.26 Da QDa Positive(+) SIR Ch2 342.26
Date Acquired: Date Processed:	2/29/2016 3:44:07 PM EST 6/14/2016 11:28:49 AM EDT		Da, CV=10





	SAMPLE	INFORMATIC	) N
Sample Name: Sample Type: Vial: Injection #: Injection Volume:	mix 2 new std Standard 1:A,2 1 2.00 ul	Acquired By: Sample Set Name: Acq. Method Set: Processing Method: Channel Name:	System sample 2 reprocess screen cannabinoids isopr CEL1 ms quant 372 QDa Ch14 372.20 Da ODa Dacifiua(L) SID Ch14
Date Acquired: Date Processed:	2/29/2016 3:44:07 PM EST 6/14/2016 11:28:49 AM EDT	Proc. Chnr. Descr	372.20 Da, CV=10



# Empower® 3

## screen cannabinoids

	SAMPLE	INFORMATIC	) N
Sample Name: Sample Type: Vial: Injection #: Injection Volume: Run Time:	mix 2 new sample 1 Unknown 1:A,3 1 2.00 ul 11.0 Minutes	Acquired By: Sample Set Name: Acq. Method Set: Processing Method: Channel Name: Proc. Chnl. Descr.:	System sample 2 reprocess screen cannabinoids isopr CEL1 lurie syn can screen Extract 210.0 PDA Spectrum (210-350)nm
Date Acquired: Date Processed:	2/29/2016 4:10:19 PM EST 6/14/2016 11:14:37 AM EDT	-	







#### PDA Result Table

	RT	Purity1 Ang le	Purity1 Threshold	Match1 Spect. Name	Match1 Angle	Match1 Threshold	PDA/FLR Match2 Spect. Name	PDA/FLR Match2 Angle	PDA/FLR Match3 Spect. Name	PDA/FLR Match3 Angle	Base Peak (m/z)
1	1.555	22.106	21.733								282.19
2	5.125	0.304	0.381	JWH 19	0.403	1.181	JWH 18	0.408	JWH 73	0.411	342.16
3	9.531	2.516	1.924	JWH 081	3.585	2.506	JWH 200	9.573	AM 694	12.613	372.23

#### PDA Result Table

	MS Match1 Spect. Name	MS Match2 Spect. Name	MS Match3 Spect. Name
1	RCS4	UR 144	XLR 11
2	JWH 18	RCS 4	UR 144
3	JWH 81	CP 47, 497	JWH 250



	SAMPLE	NFORMAT	ION
Sample Name: Sample Type: Vial: Injection #: Injection Volume: Run Time:	mix 2 new sample 1 Unknown 1:A,3 1 2.00 ul 11.0 Minutes	Acquired By: Sample Set Name: Acq. Method Set: Processing Method: Channel Name:	System sample 2 reprocess screen cannabinoids isopr CEL1 uv quant 215 comp PDA Ch2 215nm@4.8nm -Compens.
Date Acquired: Date Processed:	2/29/2016 4:10:19 PM EST 6/14/2016 11:28:51 AM EDT		





	SAMPLE	INFORMATIC	D N
Sample Name: Sample Type: Vial: Injection #: Injection Volume: Pun Time:	mix 2 new sample 1 Unknown 1:A,3 1 2.00 ul 11.0 Minutes	Acquired By: Sample Set Name: Acq. Method Set: Processing Method: Channel Name: Proc. Chal. Descr.	System sample 2 reprocess screen cannabinoids isopr CEL1 ms quant 342 QDa Ch2 342.26 Da ODa Positive(+) SIR Ch2 342 26
Date Acquired: Date Processed:	2/29/2016 4:10:19 PM EST 6/14/2016 11:28:51 AM EDT	FIUC. CHIII. Desci	Da, CV=10





	SAMPLE	INFORMATIC	) N
Sample Name: Sample Type: Vial: Injection #: Injection Volume: Run Time:	mix 2 new sample 1 Unknown 1:A,3 1 2.00 ul 11.0 Minutes	Acquired By: Sample Set Name: Acq. Method Set: Processing Method: Channel Name: Proc. Chnl. Descr.:	System sample 2 reprocess screen cannabinoids isopr CEL1 ms quant 372 QDa Ch14 372.20 Da QDa Positive(+) SIR Ch14
Date Acquired: Date Processed:	2/29/2016 4:10:19 PM EST 6/14/2016 11:28:52 AM ED	г	372.20 Da, 6V - 10



## S4 screen bath salts





#### PDA Result Table

	RT	Purity1 Angle	Purity1 Threshold	Match1 Spect. Name	Match1 Angle	Match1 Threshold	PDA/FLR Match2 Spect. Name	PDA/FLR Match2 Angle	PDA/FLR Match3 Spect. Name	PDA/FLR Match3 Angle	Base Peak (m/z)
1	1.037	9.061	90.000	lidocaine	21.067	90.000					279.06
2	1.062	13.766	90.000	lidocaine	12.163	90.000					191.03
3	1.091	13.046	90.000	lidocaine	11.178	90.000					191.00
4	1.669	0.955	8.614	Buphedrone	0.699	5.686	Pentedrone	1.071	Methcathinone	1.603	192.10
5	2.850	0.520	5.609	Mephedrone	1.064	4.156	4 Methylethcathinone	1.961	4 MePPP	5.121	178.07

#### PDA Result Table

	MS Match1 Spect. Name	MS Match2 Spect. Name	MS Match3 Spect. Name
1	4-methylethcathinone	lidocaine	MDPV
2	lidocaine	MDPV	Naphyrone
3	4-methylethcathinone	lidocaine	MDPV
4	4-methylethcathinone	Pentedrone	lidocaine
5	Mephedrone	Buphedrone	Naphyrone



### quantitation bath salts

# SAMPLE INFORMATION

Sample Name: Sample Type: Vial: Injection #: Injection Volume: Run Time: Dilution 1.0000	mix 14 standard Standard 2:E,1 1 0.50 ul 8.0 Minutes	Acquired By: Sample Set Name: Acq. Method Set: Processing Method: Channel Name:	System sample 14 UV reprocess screen bath salts bath salts quant comp 240nm PDA Ch3 240nm@4.8nm - Compens.
Date Acquired: Date Processed:	5/12/2016 5:12:49 PM EDT 6/14/2016 2:00:53 PM EDT		





## quantitation bath salts

	SAMPLE	INFORMAT	ION
Sample Name: Sample Type: Vial: Injection #: Injection Volume: Run Time: Dilution 1.0000 Date Acquired: Date Processed:	mix 14 standard Standard 2:E,1 1 0.50 ul 8.0 Minutes 5/12/2016 5:12:49 PM EDT 6/14/2016 2:00:54 PM EDT	Acquired By: Sample Set Name: Method Set: Processing Method: Channel Name:	System sample 14 UV reprocess Acq. screen bath salts bath salts quant comp 250nm PDA Ch1 250nm@4.8nm -Compens.





## quantitation MS bath salts

	SAMPLE	INFORMATIC	) N
Sample Name: Sample Type: Vial: Injection #: Dilution Run Time: Injection Volume: Date Acquired: Date Processed:	mix 14 standard dilution Standard 2:C,5 1 1.0000 8.0 Minutes 0.50 ul 5/12/2016 12:02:37 AM EDT 6/14/2016 1:58:28 PM EDT	Acquired By: Sample Set Name: Acq. Method Set: Processing Method: Channel Name: Proc. Chnl. Descr.:	System mix 14 MS reprocess screen bath salts bath salts 178 QDa Ch3 178.12 Da QDa Positive(+) SIR Ch3 178.12 Da, CV=15





## quantitation MS bath salts

	SAMPLE	INFORMATIC	) N
Sample Name: Sample Type: Vial: Injection #: Dilution Run Time: Injection Volume: Date Acquired: Date Processed:	mix 14 standard dilution Standard 2:C,5 1 1.0000 8.0 Minutes 0.50 ul 5/12/2016 12:02:37 AM EDT 6/14/2016 1:58:28 PM EDT	Acquired By: Sample Set Name: Acq. Method Set: Processing Method: Channel Name: Proc. Chnl. Descr.:	System mix 14 MS reprocess screen bath salts bath salts 192 QDa Ch5 192.14 Da QDa Positive(+) SIR Ch5 192.14 Da, CV=15



### screen bath salts





#### PDA Result Table

	RT	Purity1 Angle	Purity1 Threshold	Match1 Spect. Name	Match1 Angle	Match1 Threshold	PDA/FLR Match2 Spect. Name	PDA/FLR Match2 Angle	PDA/FLR Match3 Spect. Name	PDA/FLR Match3 Angle	Base Peak (m/z)
1	0.968	33.954	90.000	lidocaine	12.925	90.000					279.07
2	1.050	13.830	90.000	lidocaine	13.606	90.000					191.01
3	1.658	0.929	8.536	Buphedrone	0.565	5.687	Pentedrone	0.801	Methcathinone	1.343	192.09
4	2.827	0.810	5.623	Mephedrone	0.840	4.350	4 Methylethcathinone	1.763	4 MePPP	4.929	178.06

#### **PDA Result Table**

	MS Match1 Spect. Name	MS Match2 Spect. Name	MS Match3 Spect. Name
1	Butylone	lidocaine	MDPV
2	lidocaine	MDPV	Naphyrone
3	4-methylethcathinone	Pentedrone	lidocaine
4	Buphedrone	Mephedrone	lidocaine



## quantitation bath salts

	SAMPLE	INFORMAT	ION
Sample Name: Sample Type: Vial: Injection #: Injection Volume: Run Time: Dilution 1.0000 Date Acquired: Date Processed:	mix 14 sample 1 Unknown 2:E,2 1 0.50 ul 8.0 Minutes 5/12/2016 5:30:51 PM EDT 6/14/2016 2:00:57 PM EDT	Acquired By: Sample Set Name: Method Set: Processing Method: Channel Name:	System sample 14 UV reprocess Acq. screen bath salts bath salts quant comp 240nm PDA Ch3 240nm@4.8nm-Compens.





## quantitation bath salts

	SAMPLE	INFORMA	TION
Sample Name: Sample Type: Vial: Injection #: Injection Volume:	mix 14 sample 1 Unknown 2:E,2 1 0.50 ul	Acquired By: Sample Set Name: Method Set: Processing Method: Channel Name:	System sample 14 UV reprocess Acq. screen bath salts bath salts quant comp 250nm PDA Ch1 250nm@4.8nm-Compens.
Run Time: Dilution 1.0000 Date Acquired: Date Processed:	8.0 Minutes 5/12/2016 5:30:51 PM EDT 6/14/2016 2:00:59 PM EDT		





## quantitation MS bath salts

	SAMPLE	INFORMATIC	) N
Sample Name: Sample Type: Vial: Injection #: Dilution Run Time: Injection Volume: Date Acquired: Date Processed:	mix 14 sample 1 dilution Unknown 2:C,6 1 100.0000 8.0 Minutes 0.50 ul 5/12/2016 12:20:40 AM EDT 6/14/2016 1:58:29 PM EDT	Acquired By: Sample Set Name: Acq. Method Set: Processing Method: Channel Name: Proc. Chnl. Descr.:	System mix 14 MS reprocess screen bath salts bath salts 178 QDa Ch3 178.12 Da QDa Positive(+) SIR Ch3 178.12 Da, CV=15





## quantitation MS bath salts

	SAMPLE	INFORMATIC	) N
Sample Name: Sample Type: Vial: Injection #: Dilution Run Time: Injection Volume: Date Acquired: Date Processed:	mix 14 sample 1 dilution Unknown 2:C,6 1 100.0000 8.0 Minutes 0.50 ul 5/12/2016 12:20:40 AM EDT 6/14/2016 1:58:30 PM EDT	Acquired By: Sample Set Name: Acq. Method Set: Processing Method: Channel Name: Proc. Chnl. Descr.:	System mix 14 MS reprocess screen bath salts bath salts 192 QDa Ch5 192.14 Da QDa Positive(+) SIR Ch5 192.14 Da, CV=15



#### S5 UHPSFC protocol synthetic cannabinoids

Synthetic cannabinoids drug screening or confirmation and quantitation

<u>Sample preparation</u>- Accurately weigh out 100 mg of ground [1] plant material into a 250 mL volumetric flask. Dilute to volume with solvent A containing heptane/ethyl acetate/methanol (40:20:40). Vortex sample for one minute, filter sample into a 2.0 mL glass vial through a 0.22 µm PTFE syringe filter. For quantitation dilute if necessary with solvent A for sample solution to be in linear range.

<u>Standard preparation</u>- Accurately weigh standard material of target compounds into an appropriate volumetric flask and dilute to volume with solvent A so that final concentration is approximately 4  $\mu$ g/mL. Ideally, especially for quantitation by MS, the standard concentration should be within 2X of the sample concentration.

<u>UHPSFC conditions</u>¹[2]- column- Acquity UPC² Trefoil CEL2 (2.5  $\mu$ m 3.0 x 150 mm); mobile phase conditions- injection size, 2 $\mu$ L,; Initial conditions: 20% isopropanol, 80% carbon dioxide. Final conditions: 31% isopropanol, 69% carbon dioxide, 10.3 minute linear gradient, 1.0 min gradient re-equilibration; flow rate 1.25 mL/min; temperature 55°C, ABPR 2200 psi; PDA-UV and single quadropole MS detection. See Figure 1 for chromatogram of controlled synthetic cannabinoids using UV and MS detection.

<u>Linear range and LOD's-</u> See Table 9 for linear range and LOD's for both UV and MS detection.

<u>Data analysis -</u> Empower 3- The following files are available on electronic media upon request of the PI (*optimized methods* and *MS libraries* and *UV libraries* to accomplish both drug screening and quantitation for up to three drugs).

Method Set	Channel Name	Processing Method	Report Method
screen cannabinoid 210	Extract 210.0	lurie syn can screen ^{a,b}	screen cannabinoids
lurie quant	PDA Chx 215nm@4.8	lurie syn cann 031116	lurie quantitation syn
cannabinoids	nm-Compens.	215 cmpe	can
	QDA Chx [M-H] ⁺ Da	ms quant [M-H] ⁺	lurie quantitation syn
			can
	QDA Chx [M-H] ⁺ Da	ms quant [M-H] ⁺	lurie quantitation syn
			can
	QDA Chx [M-H] ⁺ Da	ms quant [M-H] ⁺	lurie quantitation syn
			can

^aUV library - Synthetic Cannabinoids Optimal

^bMS library- Synthetic Cannabinoids Optimal

For sample reports see S3.

¹ Chromatographic conditions for Water's UHPSFC instrument. These conditions may have to be tweaked for other vendors depending on the dwell volume.

#### S6 UHPSFC protocols synthetic cathinones

Bath salt drug screening or confirmation and quantitation

<u>Sample preparation</u>- Weigh out 50 mg of powdered sample into a 500 mL volumetric flask. Dilute to volume with solvent containing methanol. Vortex sample for one minute, filter sample into a 2.0 mL glass vial through a 0.22  $\mu$ m PTFE syringe filter.

<u>Standard preparation</u>- Accurately weigh standard material of target compounds into an appropriate volumetric flask and dilute to volume with methanol so that final concentration is approximately 62  $\mu$ g/mL. For MS quantitation dilute 1/100 with methanol. Ideally, especially for quantitation by MS, the standard concentration should be within 2X of the sample concentration.

<u>UHPSFC conditions¹</u>- column- Acquity UPC² Torus DIOL (1.7  $\mu$ m 3.0 x 100 mm); mobile phase conditions- injection size, 2 $\mu$ L,; Conditions: 3% methanol with ammonium formate, 97% carbon dioxide, 8 minute linear gradient, 1.0 min gradient re-equilibration; flow rate 1.25 mL/min; temperature 40°C, ABPR 2200 psi; PDA-UV and single quadropole MS detection. See Figure 1 for chromatogram of controlled bath salts using UV and MS detection.

<u>Linear range and LOD's-</u> See Table 12 for linear range and LOD's for both UV and MS detection.

<u>Data analysis -</u> Empower 3- The following files are available on electronic media upon request of the PI (*optimized methods* and *MS libraries* and *UV libraries* to accomplish both drug screening and quantitation for up to three drugs).

Method Set	Channel Name	Processing Method	Report Method
screen bath salts	PDA 230 nm	bath salts screening ^{a,b}	screen bath salts
quantitation bath salts	PDA Chx X nm@4.8 nm- Compens.	bath salts quant com X nm	quantitation bath salts
quantitation MS bath salts	QDA Chx [M-H] ⁺ Da	bath salts [M-H] ⁺	quantitation MS bath salts

^aUV library – Bath Salts Optimal

^bMS library- Bath Salts Optimal

For sample reports see S4.

¹ Chromatographic conditions for Water's UHPSFC instrument. These conditions may have to be tweaked for other vendors depending on the dwell volume.